*Journal of Basic Physical Research ISSN: 2141-8403 Prints, 2141-8411 Online*
*Vol.9, No.1 pp. 91- 100, Jan., 2019. Available online at www.jbasicphyres-unizik.org*

# TREATMENT OF PHARMACEUTICAL EFFLUENT USING SEED OF *PHOENIX DACTYLIFERA* AS A NATURAL COAGULANT

***Ifeoma Maryjane Iloamaeke and Chizaram Onyinyechi Julius**
*Department of Pure and Industrial Chemistry, Nnamdi Azikiwe University, Awka, Nigeria*
*ifeomailoamaeke@yahoo.com or im.iloamaeke@unizik.edu.ng*

## Abstract
*Phoenix dactylifera is used to treat pharmaceutical effluent by coagulation-flocculation process. Physicochemical analysis was carried out on the effluent while the coagulant was subjected to proximate analysis and characterization. Coagulation-flocculation process was analyzed as a function of pH, settling time and dosage of the coagulant. The efficiency of coagulation-flocculation process was quantified on the colour removal. The result of physicochemical analysis obtained indicated that the effluent was highly polluted hence the, need for treatment. The proximate composition of this coagulant was found to be in this order carbohydrate (72.1%)>fat(14.9%)>protein(8.051%)>ash content(1.54%)>moisture content(1.15%). The maximum colour removal efficiency is 99.86% at the dosage of 100 mg/L, settling time of 50 minutes at constant pH of 2. The scanning electron microscopy of Phoenix dactylifera before and after treatment showed that there were changes in the surface morphology of this coagulant before and after treatment. FTIR result of the coagulant after treatment showed remarkable shift in the spectra bands, some new bands were added while some were removed; hence seed of Phoenix dactylifera can serve as an alternative coagulant.*
**Keywords**: pharmaceutical, effluent, *Phoenix dactylifera,* coagulation-flocculation

## Introduction
Effluent, according to WHO, is any wastewater treated or untreated that flows out of the treatment plant or sewer. Pharmaceutical effluent is wastewater generated by pharmaceutical industry during the process of drugs manufacturing [20, 2]. Due to challenges of diseases and man's quest to solve his problems hence demands for drugs have been promoted, which led to an increase in the number of pharmaceutical industries whose methods of drugs production involve chemical synthetic routes. Sometimes, the operation demands series of complex chemical reactions which discharged pharmaceutical effluent to the environment. This pharmaceutical effluent is characterized by high organic and inorganic chemical contents [14], colours and heavy metals [24] that if left untreated before being discharged pose health risk to the environment, receiving water bodies and farmlands. For example, if effluent has high concentration of biological oxygen demand (BOD) and chemical oxygen demand (COD), when discharged into inland rivers, streams or lakes, their biological stabilization can deplete natural oxygen resources and cause septic conditions that are detrimental to aquatic species [11]. The leaching of heavy metals and some of the pollutants can mar our crops adversely that their root will pick up these heavy metals and come back to us (the consumers). Some of them destroy vital organs like liver, kidney cause cancer, neurological disorder, Alzheimer diseases etc when accumulated in body system [13].

Coagulation-flocculation is a physicochemical method of treating wastewater which is widely used because of its simplicity and cost effectiveness. With the emergence of green chemistry natural coagulant came to the forefront since they are environmentally friendly, non-toxic and biodegradable. Some of the natural coagulant that have been studied are *Moringa oleifera*, *Cicer arietinum*, and *Dolichos lablab* [8], Dragon fruit foliage [22], Okra pod [19], *Cactus opuntia* (*ficus-indica*)[25], *Descurainia sophia L.* [16], *Corchorus Olitorius* L. [3], Aloe Vera, Moringa Oleifera seeds and Cactus [6].

*Phoenix dactylifera* is called date or date palm in English. Date palm tree belongs to *Arecaceae* family (Angiosperms, monocotyledon) [5]. It is a one-seeded fruit, usually oblong as shown in Figure 1. It has been the staple food and chief source of wealth in the irrigable desert from ancient times [1]. *Phoenix dactylifera* has been found to originate from North Africa and grows in

country such as Arabia , Persian Gulf, Canary Islands, in the northern Mediterranean and in the south of the United States. It has palm with a very slender trunk, up to 30 m tall, conspicuously covered with the remains of sheaths from fallen leaves. Its leaflets are clustered together in a maximum number of 20-30 and forming a loose crownshaft. The leaves also are pinnated up to 6 m long, upper leaves are ascending, basal leaves are recurved, the segments are coriaceous, linear, rigid, sharp pointed and blue-green in colour [21].
The study is aimed at treating pharmaceutical effluent with the seed of *Phoenix dactylifera* in order to remove colour from the effluent.



**Figure 1: Phoenix dactylifera fruit**



**Figure 2: The seed of *Phoenix dactylifera***

## Materials and methods

### Collection of pharmaceutical effluent samples

The pharmaceutical effluent was collected from the waste channel of a pharmaceutical industry at Awka, Anambra, state, Nigeria. The colour of this effluent is purple. The effluent was collected in a 10 litres polyethylene bottle tightly closed and stored in a cool dry place.

### Preparation of Coagulant

*Phoenix dactylifera* was purchased at Eke Awka market, Awka, Anambra State, the edible body was eaten leaving the seed which was sun-dried for two weeks to remove moisture after which it was ground using a grinder until its particles were reduced to powder of particle size 1.18 $\mu m$ using sieve, after which it was kept in a cool, dry container.

### Physicochemical analysis

Determination of chemical oxygen demand (COD), biological oxygen demand (BOD), total dissolved solid (TDS), total suspended solid (TSS), total hardness, chloride, phosphate, nitrate and sulphate were carried out according to the standard method for the examination of water and wastewater [9, 7]. Electrical conductivity, turbidity and pH were determined using electrical conductivity meter Model DDS-307, Turbid meter Lab. Tech. model 038, pH meter Hanna model HI991300.

### Characterization of the *Phoenix dactylifera* seed

The seed of *Phoenix dactylifera* was characterized using standard methods and instruments [23].

### Coagulation-Flocculation Experiment

The initial absorbance of the pharmaceutical effluent was taken at 510 nm. Exactly 100 mg of the coagulant (*Phoenix dactylifera*) was weighed into 250 mL beaker and 100 mL of the effluent which was made to be at pH 2 was added into it. Desired pH was obtained by adding (1M HCl for acidic pH or 1M NaOH for basic pH) to the effluent. The content of the beaker was stirred for 2 mins of rapid mixing (100 rpm) and 20 mins for slow mixing (40 rpm) with a magnetic stirrer and

then left undisturbed for 50 mins. It was filtered and the filtrate was subjected to colour removal analysis using UV/VIS spectrophotometer at 510 nm as $\lambda_{max}$. The experiment was repeated for the dosages of 200-500 mg/L, pH 4, 6,8,10, then time of 10-50 mins and dosage of 100-500 mg/L at constant pH 2. The colour removal efficiency is calculated thus:

$$\%\ \text{colour removal efficiency} = \frac{\text{Initial absorbance} - \text{finial absorbance}}{\text{Initial absorbance}} \times 100\%$$

**Results and discussions**

**Physicochemical analysis of the pharmaceutical effluent**

The result of the physicochemical analysis of the pharmaceutical effluent is shown in Table 1. It can be inferred from the result that all the physicochemical characteristics were within WHO limit of effluent discharge, except total hardness, and turbidity. This pharmaceutical effluent is observed to be purple with absorbance of 5.073 hence the need for treatment for the removal of colour before being discharged into the environment. The presence of colour in effluent is caused by various manufacturing processes and the synthetic chemical dyes. Colour affects the aesthetic value of water, if coloured effluent is discharged to nature, photosynthesis activity is limited which will have prejudicial effects on aquatic ecosystems [3].

**Table 1: Physicochemical characteristic of the pharmaceutical Effluent**

| Parameters | Concentration | WHO (STD) | NESREA(STD) |
|---|---|---|---|
| Colour of the effluent | Purple (Absorbance of 5.073) | - | - |
| pH | 7.55 | 6-9.5 | 6-9 |
| Total hardness mg/L | 444 | 70 | - |
| Conductivity $\mu$S/cm | 359 | - | |
| Nitrate mg/L | 19.7 | 20 | 10 |
| Phosphate mg/L | 5.38 | - | 5 |
| Chloride mg/L | 172 | 250 | 600 |
| Sulphate mg/L | 431 | 500 | 500 |
| Alkalinity mg/L | 35 | - | - |
| BOD | 46.6 | - | - |
| COD | 229 | 50 | |
| TSS | 2.34 | 30 | |
| TDS | 4.18 | <1200 | 2000 |
| Turbidity (NTU) | 870 | 5 | - |

**WHO standard from Olaitan *et al*., 2014 and National Environmental Standard and Regulation Enforcement Agency of Nigeria (NESREA), 1991)**

**Table 2: Proximate composition of theseed of *Phoenix dactylifera***

| Parameters | Concentration (%) |
|---|---|
| Ash | 1.55 |
| Moisture | 1.15 |
| Fibre | 2.25 |
| Fat | 14.9 |
| Protein | 8.05 |
| Carbohydrate | 72.1 |

**Proximate analysis of the seed of *Phoenix dactylifera***

Result obtained from proximate analysis is recorded in Table 2. From the Table, the ash content of the seed of *Phoenix dactylifera* is recorded as 1.547%. The ash content shows the presence of carbon compounds and inorganic components in the form of salts and oxides [23]. Carbon plays a critical role in the adsorption and removal of metals, colour and particles from wastewater because of its porous nature [14]. This is an indication that the seed of *Phoenix dactylifera* in its granular or ash form can act as a coagulant. Moisture content is 1.15%, this shows that *Phoenix dactylifera* seed could be stored for a very long time since high percentage of moisture on seeds could lead to high microbial activities during storage hence facilitating spoilage [4]. Fibre content, fat content and protein were found to be (2.25, 14.935 and 8.05 %) respectively. Carbohydrate content of *Phoenix dactylifera* seed is 72.069%. Carbohydrate has oxygen and hydrogen elements as some of its chemical composition, therefore, the propensity of the formation of charges such as hydrogen and oxygen ions [17]. Carbohydrate has oxygen atom, in solution it becomes oxygen ion which is negatively charge and can attract metallic ions and possibly remove them from solution.

**Impact of coagulant dosage, pH and time on the colour removal efficiency**

Figures 3 and 4 are Plots of colour removal efficiency against dosage at varying pH and dosages at constant time of 50 minutes. The Figures depict that the coagulant dosages and pH have significant effect in the colour removal efficiency, thus as the coagulant dosage decreases from 500-100 mg/L, there is an increase in the colour removal efficiency of the pharmaceutical effluent. This is because above 100 mg/L of coagulant dosage, excess coagulant has been introduced, therefore, restabilization of charges take place consequently a decrease in the colour removal efficiency is detected [12,6,18]. Lowest pH favour the highest colour removal efficiency. The maximum colour removal efficiency lies at 93.81% at coagulant dosage of 100 mg/L and pH of 2 in both Figures.

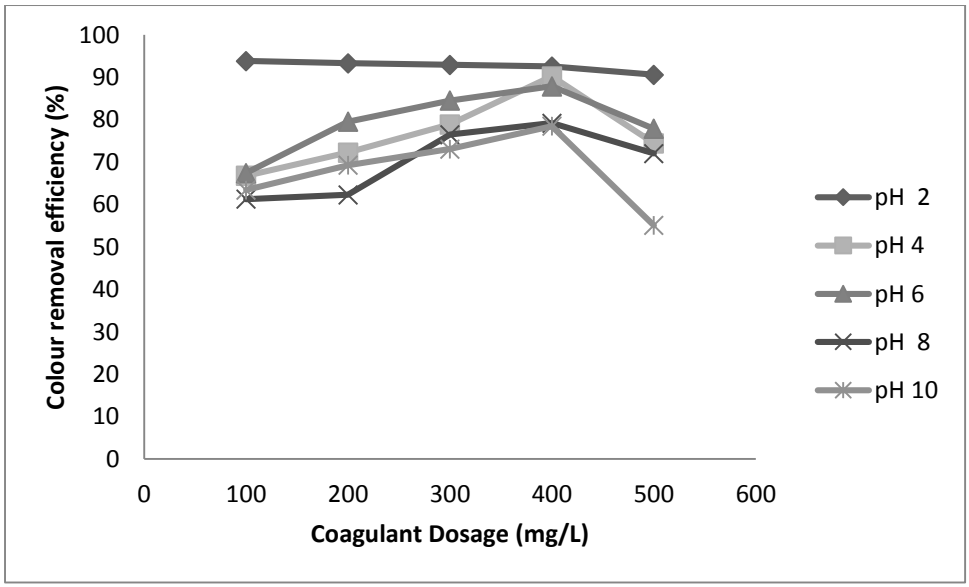



**Figure 3: Plot of colour removal efficiency against coagulant dosages with varying pH at time of 50 mins**

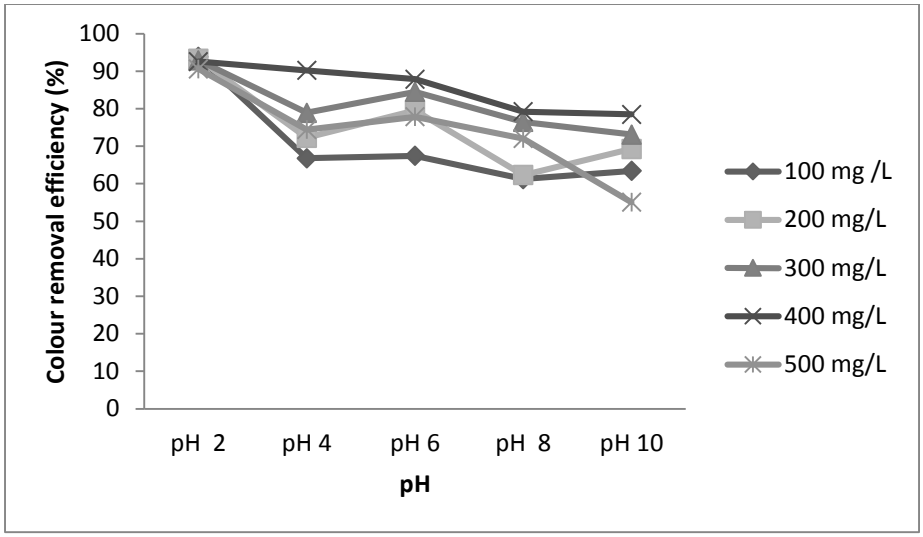



**Figure 4: Plot of % colour removal efficiency against pH with varying coagulant dosages at time of 50 mins**

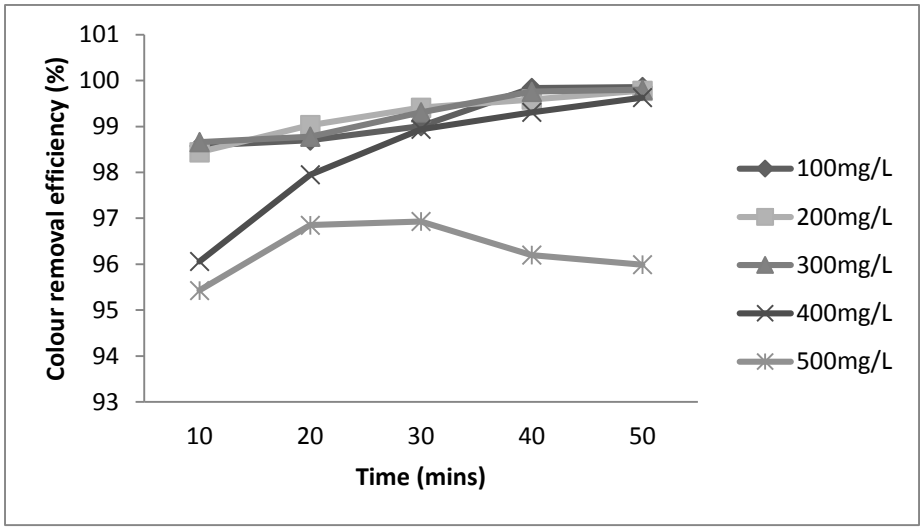



**Figure 5: Plot of % colour removal efficiency against time with varying coagulant dosages at pH 2**

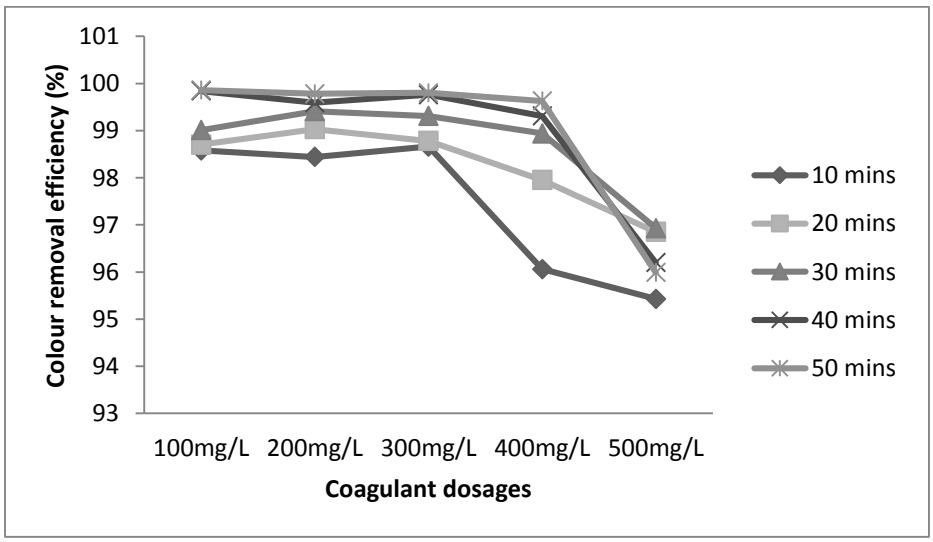



**Figure 6: Plot of % colour removal efficiency against coagulant dosages with varying time at pH 2**

Time is another significant variable that determines colour removal efficiency as can be observed in Figures 5 and 6. The observable pattern of coag-flocculation behavior in Figures 5 and 6 shows that colour removal efficiency increases as the time ascends to 50 mins with the coagulant dosage decreasing from 500 mg/l to 100 mg/L. The optimal colour removal efficiency is 99.8% obtained at the settling time of 50 mins and dosage of 100 mg/L.

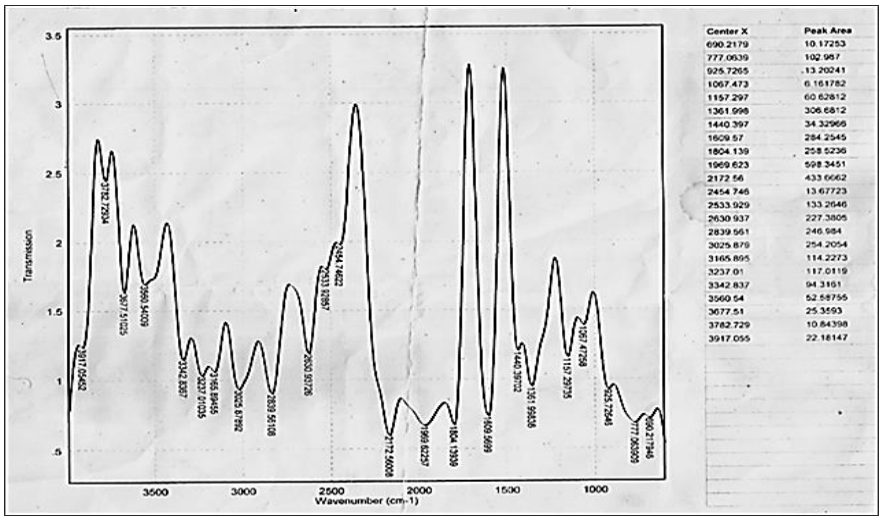



**Figure 7: FTIR spectra of Phoenix dactylifera before coagulation- flocculation process**

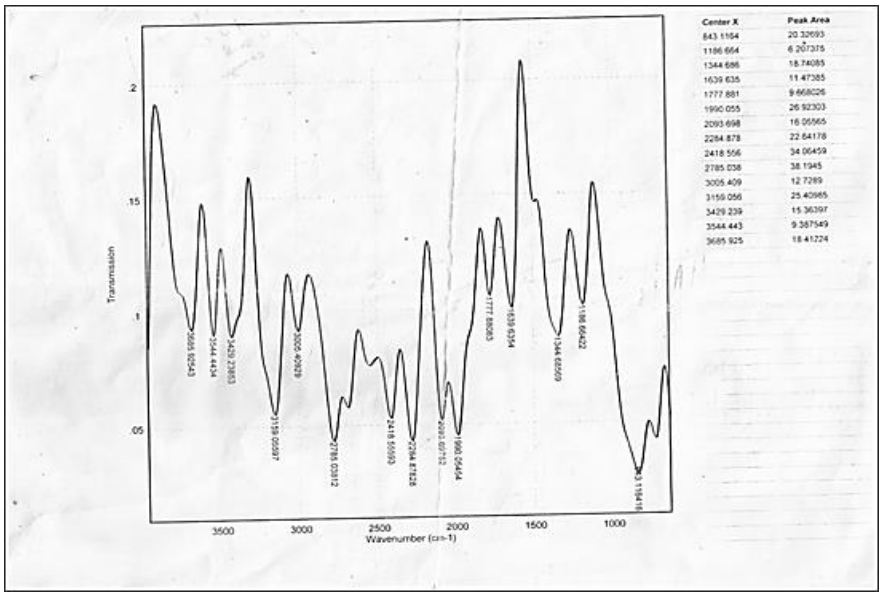



**Figure 8: FTIR spectra of Phoenix dactylifera after coagulation- flocculation process**

**IR analysis**

IR analysis of the seed of *Phoenix dactylifera* before and after coagulation-flocculation process was done using Fourier transformed infrared spectroscopy (FTIR) and the result in Figures 7 and 8. Figure 7 shows FTIR spectra of *Phoenix dactylifera* before coagulation-flocculation process. Spectra bands at 3917.055 $cm^{-1}$, 3782.729 $cm^{-1}$, 3560.540 $cm^{-1}$, 3342.837 $cm^{-1}$, 3237.010 $cm^{-1}$ and 3165.895 $cm^{-1}$ are assigned to O-H stretch which depicts the presence of alcohol. 3025.879 and 2839.561 $cm^{-1}$ denote C-H stretch from alkane group, this shows the presence of carbon and hydrogen in the date seeds. This report is in agreement with the work of Praveen, (2012) [25]. Absorption band at 2533.929 $cm^{-1}$ is O-H stretch from carboxylic acid, 2172.560 $cm^{-1}$ shows S-C≡N stretch from thiocyanate. Spectral bands at 1969.623 $cm^{-1}$, 1804.139 $cm^{-1}$ and 1609.570 $cm^{-}$

[1] are assigned C-H bend which suggests the presence of an aromatic ring. 1440.397 $cm^{-1}$ and 1361.998 $cm^{-1}$ signified O-H bend from carboxylic acid. 1157.297 $cm^{-1}$ and 1067.473 $cm^{-1}$ is C-O stretch. Absorption bands at 777.0639 $cm^{-1}$ and 690.2179 $cm^{-1}$ are C-H bend from aromatic compound.

Figure 8 is FTIR spectra of *Phoenix dactylifera* after Coagulation- flocculation process. It was observed from the Figure that the spectral bands at 3005.409 $cm^{-1}$, 3685.925 $cm^{-1}$, 3544.443 $cm^{-1}$, 3429.239$cm^{-1}$, 3159.056 $cm^{-}$1, 3005.409 $cm^{-1}$, and 2785.038 $cm^{-1}$ denote O-H stretch. 2093.698 $cm^{-1}$ indicates N=C=S stretch, 1990.055 $cm^{-1}$ and 1639.635 $cm^{-1}$ is C-H bend from aromatic ring, 1777.881$cm^{-1}$ is C=O stretch , 1344.686 $cm^{-1}$ is O-H bend, 1186.664 $cm^{-1}$ is C-O stretch. Comparing the FTIR spectra of the coagulant before and after coagulation-flocculation process (Figures 7 and 8), it can be seen that there is a remarkable shift in the spectra bands, some new bands were formed and some bands were removed in Figure 8, this is as result of interaction between the colloid particles of pharmaceutical effluent and that of the coagulant.

**SEM analysis of Phoenix *dactylifera***

Figures 9 and 10 are the SEM analyses result of the Phoenix *dactylifera* before and after coagulation-flocculation process. Critical examination of Figure 9 discloses that the surface of this coagulant look sticky and rough with tiny pores in it. The rough sticky surface and pores are the point of attachment of the colloidal particles of the pharmaceutical effluent. This property makes *Phoenix dactylifera* to be qualified as suitable coagulant.

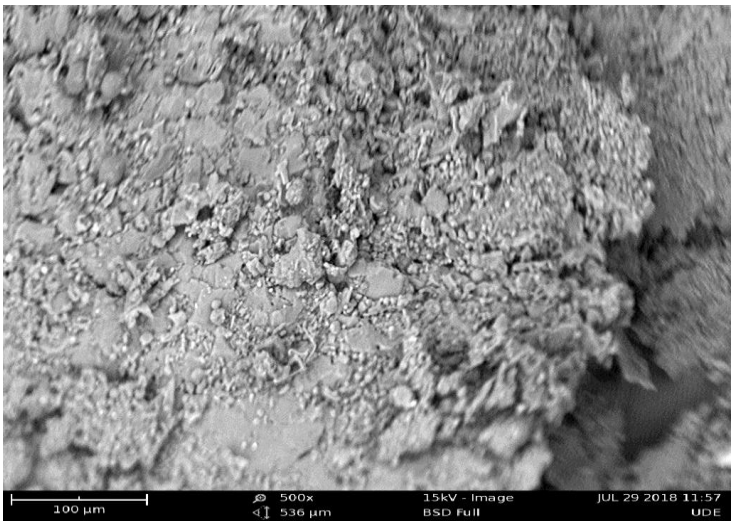



**Figure 9: SEM of Phoenix dactylifera before Coagulation- flocculation process**

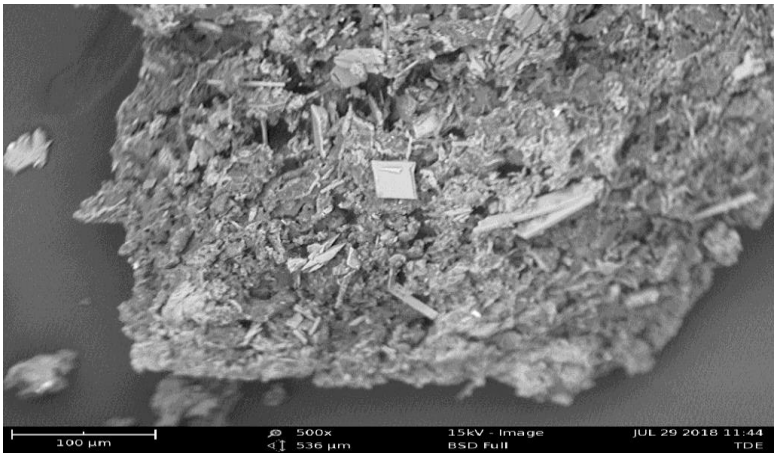



**Figure 10: SEM of Phoenix dactylifera after Coagulation- flocculation process**

Inspection of Figure 10 shows that coagulant surface is compressed as a result of the rough sticky nature of the coagulant which attracted the colloidal particles of the pharmaceutical effluent thus clumped together. These changes in the surface condition of the coagulant after coagulation-flocculation process demonstrated that the seed of *Phoenix dactylifera* removed pollutants from the pharmaceutical effluent.

**Conclusion**

At optimum condition, *Phoenix dactylifera* is found to be an effective natural coagulant in the treatment of Pharmaceutical effluent. Coagulant dosage of 100 mg/L, pH 2 and time of 50 mims were found to give the highest colour removal efficiency. FTIR spectra of the coagulant after coagulation-flocculation process indicated some shift in wave number indicating bond breakage and displacement of functional group hence color removal and other pollutants was suggested. SEM image before coagulation- flocculation process established that *Phoenix dactylifera* has rough surface and tiny pores. SEM image of the *Phoenix dactylifera* after using it to treat pharmaceutical effluent clearly showed that some particles of Pharmaceutical effluent had adsorbed on the surface of the coagulant, this further proved that the *Phoenix dactylifera* is a good coagulant for colour removal.

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